

Guidelines for Collection, Storage and transportation of Human Clinical samples for Laboratory Diagnosis of Influenza



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Laboratory confirmation of influenza

A <u>confirmed case</u> of influenza virus infection is defined as a person with an acute febrile respiratory illness with laboratory confirmed influenza virus infection by one or more of the following tests:

- Real Time PCR
- Viral culture(only in the reference laboratory)
- Four-fold rise in influenza virus specific neutralizing antibodies(only in the reference laboratory)

Clinical sample to be collected:

- A variety of Respiratory specimens can be collected including throat swab (oropharyngeal swab), Nasal swab, nasopharyngeal swabs, Bronchoalveolar lavage, tracheal aspirates, nasopharyngeal or oropharyngeal aspirates as washes.
- In case of lower respiratory tract infection (pneumonia), and intubated patients,
 Bronchoalveolar lavage or tracheal aspirates should be collected.
- Swab specimens should be collected only on swabs with a synthetic tip (such as polyester/Dacron/rayon/flocked Nylon swab) and mounted on aluminum or plastic shaft. Swabs with cotton and wooden shafts are not recommended. Specimens collected with swabs made of calcium alginate are not acceptable.
- Bronchoalveolar lavage or tracheal aspirates should be collected in a sterile screw capped container in a sufficient quantity of 3-5 ml. Alternatively, the sample collected in mucus extractor can also be sent.
- Sputum is not a preferred sample for influenza testing.

Time of Sample Collection

- As soon as possible preferably within 48 hours of onset of symptoms
- Preferably before antiviral medications are administered

Bio-safety measures for sample collection

 Before initiating collection of sample appropriate PPE should be worn by the sample collector

Personal Protective Equipment

- Masks (N-95)/ triple layered surgical masks
- Gloves
- Protective eye wear (goggles) (In case there is possibility of splash)
- Protective clothing (gown or apron)

Collection, Storage and Transport of Samples:

A. Methods of Collection

- Throat swab and Nasal swab
- Nasopharyngeal swab

Throat Swab

- Easiest sample to be collected
- Have the patient open his/her mouth wide open.
- The patient should try to resist gagging and closing the mouth during sample collection
- The posterior pharyngeal wall and tonsils should be swabbed

Nasal Swab:

- Insert swab into nostril along the lower floor of the nose till one reaches below the anterior turbinate.
- Leave the swab in place for a few seconds.
- Slowly remove swab while slightly rotating it. Use a different swab for the other nostril.
- Put tip of swab into vial containing VTM, breaking applicator's stick.

Both Nasal and Throat swabs can be collected into the same VTM to increase the viral yield.

Nasopharyngeal Swab:

This can be collected by two methods:

- Through the nasal route and
- Through oral route

Nasopharyngeal swab through Nasal route:

- Insert a thin flexible swab into nostril and right upto nasopharynx.
- Leave the swab in place for a few seconds.
- Slowly remove swab while slightly rotating it.
- Use a different swab for the other nostril.
- Put swab with tip downwards into vial containing VTM, breaking applicator's stick.
- Both the swabs can be put in same VTM vial

Nasopharyngeal swab through Oral route:

- Insert a **thin flexible swab** through mouth over the tongue and turn the swab upwards behind the soft palate to reach the nasopharynx.
- Leave the swab in place for a few seconds.





• Slowly remove swab and put the swab with tip downwards into vial containing VTM,

swab stick.

B. Labelling the Samples

Preferably use pre-printed barcode*
 labels.

breaking the extra portion of the

 In case of non-availability, the following label can be pasted on the specimen container. <u>Label</u>

Patient ID No.:

Patient's Name:

Hospital Name:

Date of sample collection:

- The label material should be made of leucoplast available in the hospital.
- ONLY ball point pen to be used for labeling (don't use ink /gel pen).
- The sample should always be accompanied with clinical proforma duly filled by the physician.

C. Storage of samples:

- Sample should be sent to the laboratory as soon as possible at 4 -8°C but not later than 48 hours
- In case of delay beyond 48 hours, the specimens must be stored at -70 °C.
- · Avoid repeated freezing and thawing of the samples

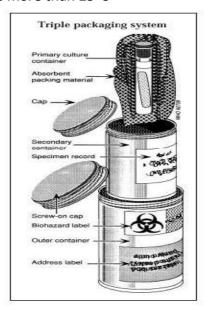
D. Transportation of samples

<u>Transportation of sample from OPD/Ward/Casuality/ICU to the laboratory within the same institute</u>

- Tightly closed and appropriately labeled VTM vial containing the sample should be transported to lab in a closed box (preferably vaccine carrier) with absorbent material (cotton /tissue paper) wrapped around the vial immediately after collection.
- Sample should be preferably transported in cold chain (4-8°C).
- The sample should never be exposed to temperature more than 25°C

<u>Transportation of sample from the Hospital/Institution to the laboratory located elsewhere</u>

- All samples should be transported after proper packaging using the standard triple packaging system https://www.who.int/csr/emc97 3.pdf
- Absorbent cotton, tissue paper, waste newspaper for wrapping primary container.
 Secondary container to hold primary container i.e. bigger tube or sealed plastic bag. Insulated ice box



with icepack, sample proforma fastened onto the secondary container.

- Sample should accompany the clinical details as per proforma enclosed in Annexure
- While transportation cold chain should be maintained.

Waste Disposal: should be done as per guidelines of your hospital

PATIENT PROFORMA FOR INFLUENZA TESTING

Patient's Name	CR/0	OPD No		
AgeSex	Tel. No)		
Address		Block		
District	State			
Occupation	Date of o	nset of illness	· ···	
Name of hospital				
District	State			
Name of Doctor/Health p	personal			
Tel.:	E mail ID		. 	
Clinical Signs & sympton	ns (encircle all that	are present in the patient):	
of breath/ Difficulty in b (in children)/ Refusal to a Pre-existing medical con	oreathing/ Hemopt accept feeds (in chil ditions (encircle all	ysis/ Cyanosis/ Hypotensic dren)/ Irritability (in childre I that are present in the pa	tient):	
that impair breathing specifyimmunocompromised/ P	or clearance of / renal dy regnancy – (If Yes,	respiratory secretions/ of sfunction/ haemoglobinory Mention Trimester)	ronic neurological conditions chronic metabolic diseases, pathies/ immunosuppressed/ /children 6 months – 18 years	
Data of sample collection	1			
Sample collected: throat swab/nasopharyngeal swab/other				
No. of samples collected				
History of Vaccination: Yes/No, If Yes then, date of vaccination:				
Treatment History:				
Antiviral Treatment taker	n:	Yes	No	
Drug	Date Initiated	Date Discontinued	Dosage (If known)	
Oseltamivir				
Other (Specify)				
Investigations Done:		Yes	No	
Chest X-Ray findings				
Outcome				
			Signature	

Name:

Designation:

For Laboratory Use only:	
Name of the laboratory:	
Date of sample received:	
Date of result:	
Lab result of influenza testing	
	Signature of Lab in charge
	Name:
	Designation: